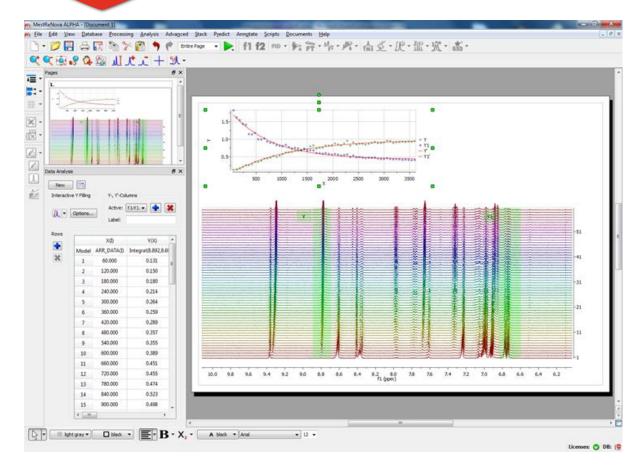


for Reaction Monitoring by NMR



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Version 8.0 Aug. 2012

Mnova

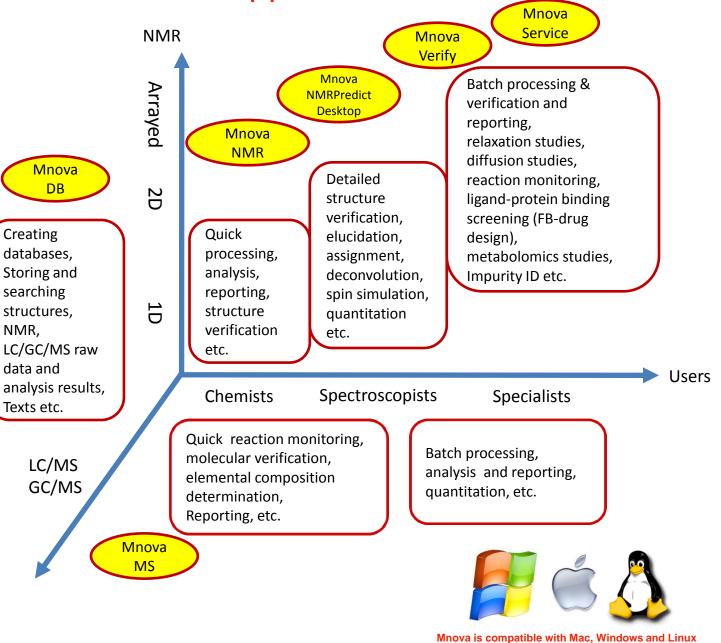


Outline

- About Mestrelab Research
- Importing and processing arrayed NMR spectra
- Extracting spectral information
- Fitting spectral data to a kinetics function
- Summary



Products and Applications





Mestrelab Research

- 1996: A research project in University of Santiago de Compostela, Spain, developed free MestReC software for NMR processing
- ▶ 2004: Mestrelab Research incorporated in Santiago de Compostela
- ▶ 2004: New MestreNova (Mnova) platform and NMR plugin released
- 2006: NMRPredict Desktop plugin released with Modgraph
- ▶ 2009: LC/GC/MS plugin released with Sierra Analytics
- 2009: Global Spectral Deconvolution (GSD) algorithm released with ExtraByte
- 2011: **DB** plugin for Database Management
- 2012: Verify plugin for auto structure verification and peak assignment
- 2012: qNMR and Screen plugins to be released
- An R&D company with >20 people and >80,000 registered users

www.mestrelab.com

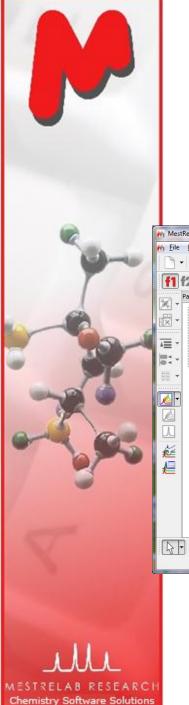


To open and process reaction monitoring data

- In a typical kinetics or reaction monitoring (RM) experiment, a series of spectra are recorded at predetermined time intervals to follow the progress of the reaction:
 - All spectra are acquired and stored as *individual spectra*
 - All spectra are acquired and stored as a single NMR experiment in *arrayed mode*
- Mnova supports both kinds of data
- For RM data acquired on individual basis: Run the Directory Spectra Stack script to open and stack all spectra under a base directory:

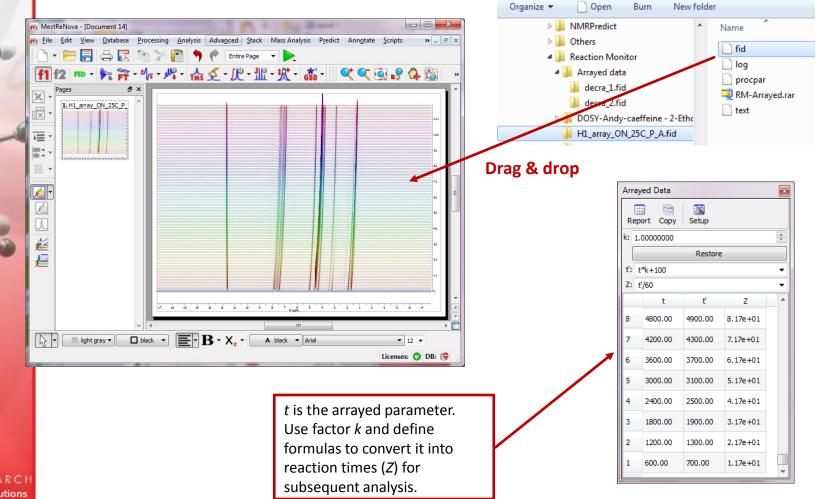
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ø	Run Script			
	Recent Scripts	•	E	
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Expo	Export Bruker Time Corrected FID		ι.	
	Import	•	ACDL	ACDLabs 1D NMR
	Report	•	Dire	Directory Spectra Stack
	NMR Analysis	•	Int3	Int32 WIN-NMR
_			LCHo	LCModel ASCII
			Phil	Philips Achieva
			Siem	Siemens Syngo
			Mult	Multi-Open

- The user selects the directory where the spectra are located
- Mnova opens and process all those spectra (FIDs) automatically using the processing parameters from the instrument
- Mnova stacks all spectra in the alphabetic order of the folder name of the experiments (You can change the order later – see subsequent slides)



To open and process reaction monitoring data

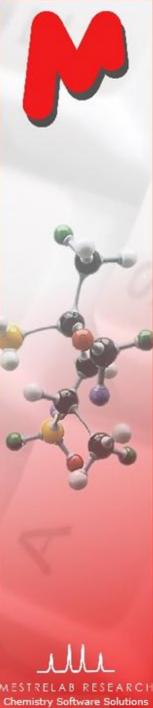
- For RM data acquired in arrayed mode: Just open the FID (Varian) or SER file (Bruker), and the individual spectra will be processed and stacked.
 - Mnova parses the arrayed parameters automatically and displays them in the Arrayed Data Table (Choose View | Tables | Arrayed Data to open it)



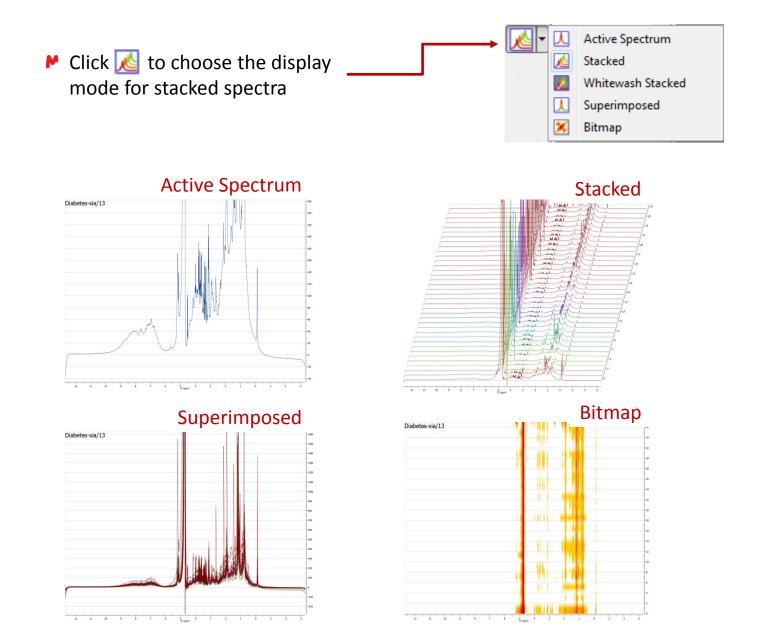


Easy handling multiple spectra in Mnova

- With Mnova, it is very easy to
 - To display stacked spectra in different modes, such as Stacked, Superimposed, Active Spectrum, or Bitmap
 - To re-process all the spectra together, such as phasing and baseline correction (this is the default)
 - To select one or several spectra and apply processing only to them
 - To hide any number of spectra for better visualization
 - To correct global or local spectral misalignment
 - To normalize the Y-scale of the spectra in various ways



To change the stacking mode





To re-process the stacked spectra

- Click 2 to toggle on the Stacked Spectra Table
- Use this table to do the following:
 - Delete spectra from the stack
 - Change order of the spectra in the stack
 - Change the Y-intensity of selected spectra
 - Change which ones to display
 - Change which ones to re-process, such as phasing, baseline correction etc.

Click and drag here to change the order of a spectrum in the stack

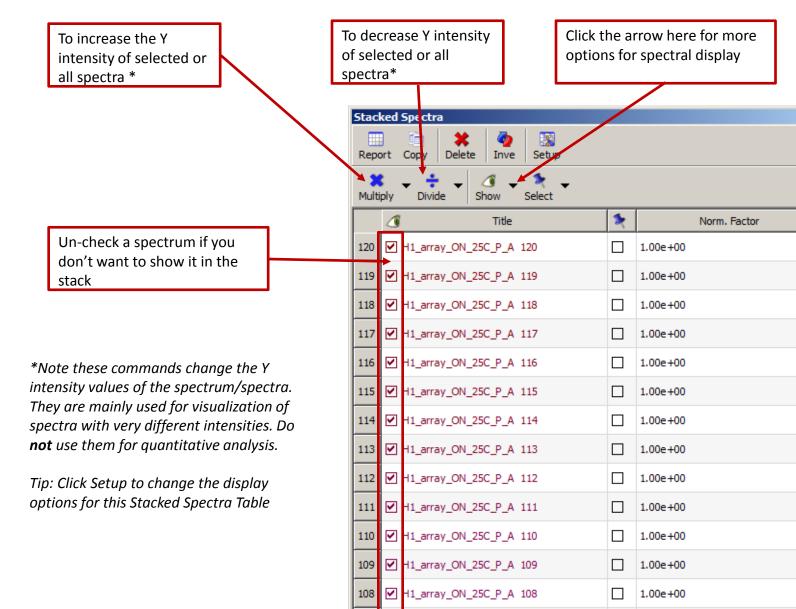
Tip: The Titles of the spectra can be customized by double-clicking on the stacked spectra, and changing the Titlerelated settings in the Properties dialog. For arrayed spectra, use "Spectrum Number" as the title or part of the title.

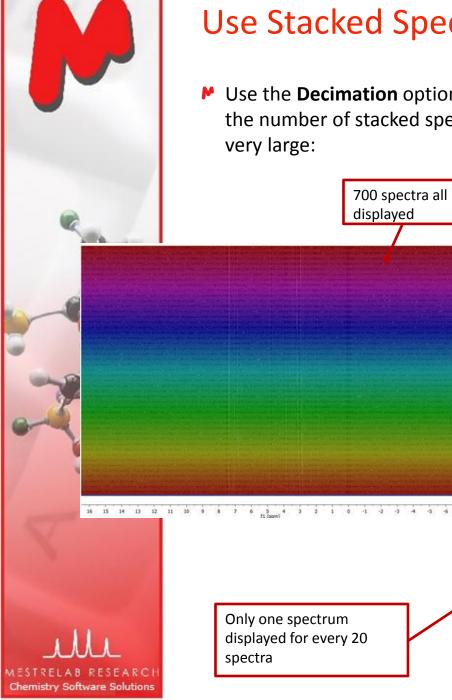
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Sho	* * ` *				
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8	Diabetes-xia/8		1.47e+04	1.00e+00	1.00e+00
7	Diabetes-xia/7		1.41e+04	1.00e+00	1.00e+00
6	Diabetes-xia/6		1.34e+04	1.00e+00	1.00e+00
5	Diabetes-xia/5		5.57e+03	1.00e+00	1.00e+00
4	Diabetes-xia/4		4.80e+03	1.00e+00	1.00e+00
3	Diabetes-xia/3		3.98e+03	1.00e+00	1.00e+00
2	Diabetes-xia/2		2.32e+03	1.00e+00	1.00e+00
1	Diabetes-xia/1		0.00e+00	1.00e+00	1.00e+00

ž



Use Stacked Spectra Table to setup the display





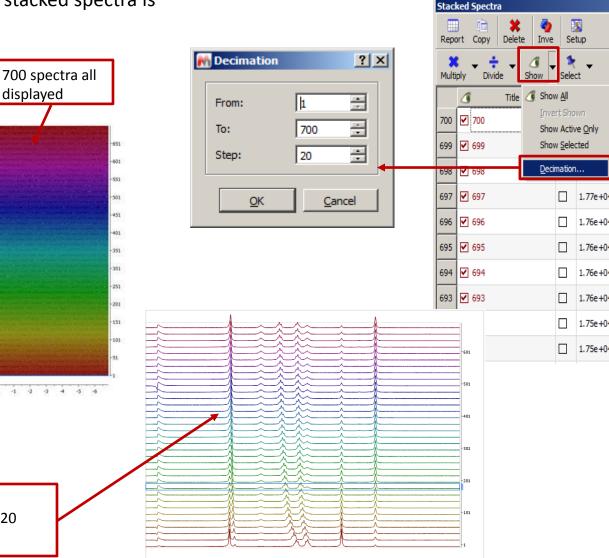
Use Stacked Spectra Table to setup the display

Use the **Decimation** option when M the number of stacked spectra is very large:

Only one spectrum displayed for every 20

spectra

displayed



4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 1.6 1.4 1.2 1.0



To re-process all or selected spectra

Spectra are automatically processed when they are opened, but sometimes you need to manually re-process all or some of them

Which ones will be changed when you apply the processing tools?

- If no spectrum is checked in the Select column, all spectra will be changed
- If some spectra are checked in the Select column, only the selected ones will be changed
- Use Undo/Redo if you've made a mistake

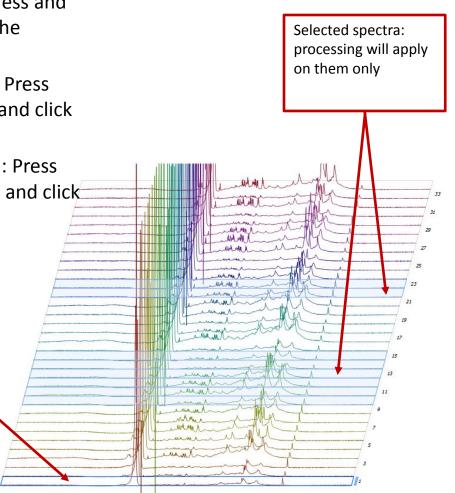
mor	re options to	Check here to choose which spectra to change				
	ked Spectra					
Mult	ply Divide Show Select					
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120	✓ H1_array_ON_25C_P_A 120		1.00e+00			
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115	✓ H1_array_ON_25C_P_A 115		1.00e+00			
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111	H1_array_ON_25C_P_A 111		1.00e+00			
110	✓ H1_array_ON_25C_P_A 110		1.00e+00			
109	✓ H1_array_ON_25C_P_A 109		1.00e+00			



To select spectra from the stacked spectra

- Spectra can also be selected from the spectral stack directly.
- To select one spectrum: Press and hold Alt key, and click on the spectrum
- To select multiple spectra: Press and hold Ctrl or Shift key, and click on a spectrum
- To de-select one spectrum: Press and hold Ctrl and Alt keys, and click on the spectrum

The active spectrum: It will be displayed when in *Active spectrum* mode. It is also used as reference spectrum in some operations such as spectral alignment





To correct phase errors and baseline

- Click for phase correction if peaks are not symmetric. Options:
 - Global method for all positive peaks
 - Selective method for positive/negative peaks
 - Metabolomics method when there is residual solvent peaks
 - Whitening method by maximizing the baseline regions
 - You can combine any of the methods listed above
 - Manual method if none of the above works

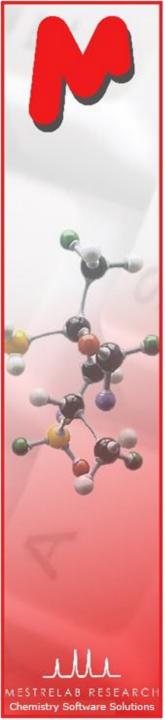
Click ^{*} for baseline correction if baseline is not zero. Options:

- Polynomial Fit
- Bernstein Polynomial Fit
- Whittaker Smoother
- Manual 🕨

Automatic	Phase Correction Options
Global	☐ Selective ☐ Metabonomics ☐ Whitening
Initial Phase:	Imported 💌
	OK Cancel

Phase Correction	×
f2 f1	
Click here and drag mouse up or down holding: left button for PH0 correction or right button for PH1 correction. (hold Ctrl key for fine tune)	
PH0: -165.02 +180 PH1: 19.26 +	
Pivot Point	1
Position: 4.072 Biggest	

Baseline Correction along f1	? ×
Method	Ok
Polynomial Fit 👻	Cancel
Whittaker Smoother Polynomial Fit	rec Extract
Bernstein Polynomial Fit	
Filter (23.93 Hz) 143 🛨 🔽	Autodetect
<u> </u>	I I I
Polynomial Order: 3	

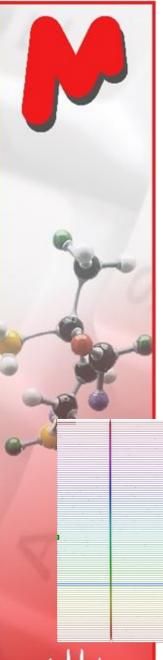


To align spectra by correcting reference

- Systematic errors of chemical shift reference can be corrected if there is an internal reference peak, e.g. TMS peak.
- Click 1 and then click on the reference peak in the active spectrum
- In the following dialog, set the proper chemical shift for the reference peak, check Auto Tune, and define a tuning region (e.g. +/- 0.05 ppm) *:

Reference along f1	
Old Shift: -0.128 ppm New Shift: 0.000 ppm Annotation	
OK Cancel Solven	
	0.10 0.06 0.02 -0.02 f1 (ppm) -0.08
	Old Shift: -0.128 ppm Image: New Shift: 0.000 ppm Image: New Shift: 0.000 ppm Image: New Shift: 0.000 ppm

*Note: it is important to make sure that the reference peak is always the highest one within the tuning region for all spectra



To correct local peak misalignment

- Zoom into the region of interest, select Advanced | Align Spectra.
- Click Item, then click-and-drag to cover the peaks to align. Click Preview to see the alignment result. Adjust other parameters until satisfactory.
- Move to other regions to continue this process until done.
- Click OK to accept the results

✓ Apply Alignment ▲ Apply Global Alignment ✓ Use Derivative Aligning Regions: ✓ ● ● ● ● ● ● ● ● ● ● ● ● ● ● ● ● ● ● ●	Image: Apply Global Alignment Image: Use Derivative Aligning Regions: Image: Prometry To	Apply Global Alignment Use Derivative Aligning Regions: Image: Comparison of the second seco	Apply Global Alignment Use Derivative Aligning Regions: Image: Comparison of the second s	Aligning Regions:	1	lign Spectra	×				
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	QK <u>Cancel</u> <u>Preview</u>	OK Cancel Preview					•				
QK <u>Cancel</u> <u>Preview</u>											
OK Cancel Preview			-11	-11	-1						



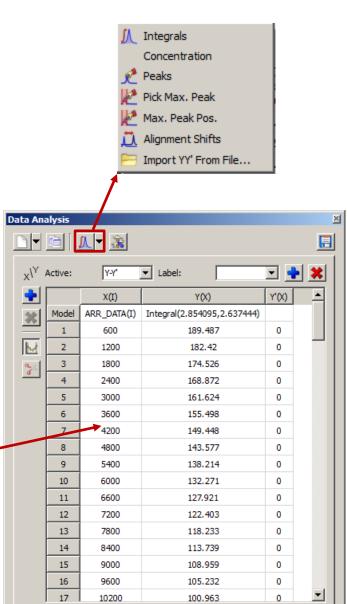
Tip: When there is peak cross-over, it may not be good idea to use local peak alignment. Instead, use the UI feature to change the integration regions so that they follow the change of the peak locations. See subsequent slides.



To analyze stacked spectra using the Data Analysis Panel

- The Data Analysis Panel provides an intuitive way to analyze multiple stacked spectral data
- Choose View | Panels | Data Analysis to open the Data Analysis Panel
- Click Create Empty Graph Tool 1 to create a new analysis.
- Choose one of the data extraction modes (e.g. Integrals), click and drag in the spectra to define the range for extracting such information (e.g. integrating the peaks).

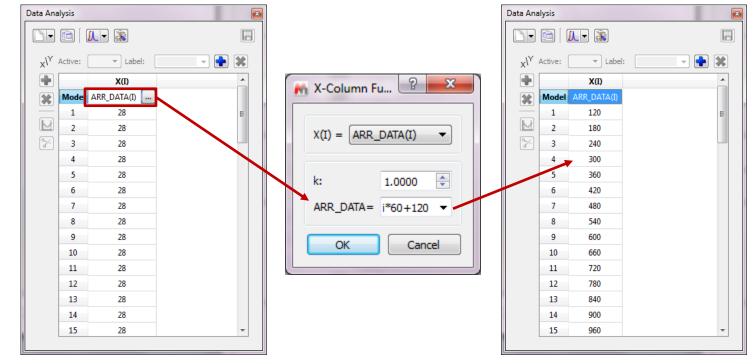
The X(I) column is automatically filled with the reaction time. Use Arrayed Data Table to preview and convert those data. You can also manually edit these data, or copy from a .txt file.

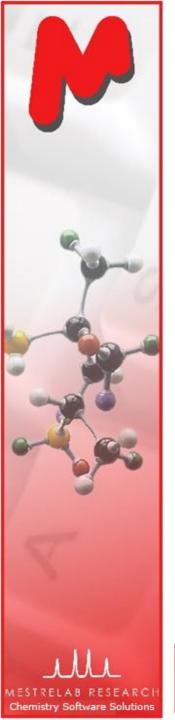




What if reaction time is not properly read?

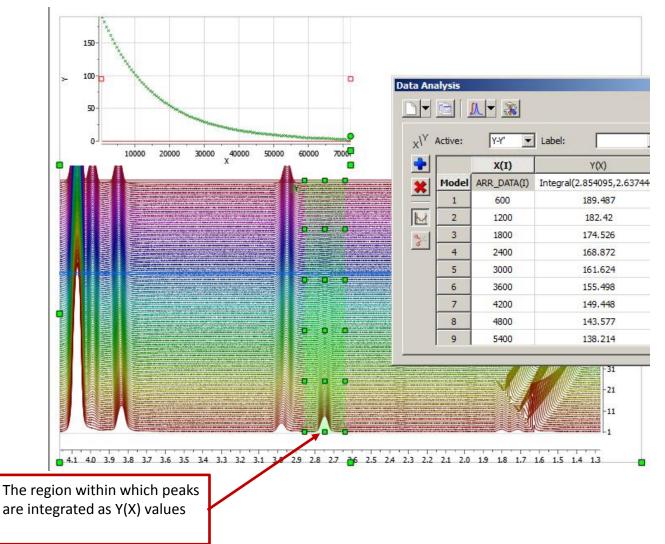
- Normally the reaction times are automatically read to Arrayed Data Table (see previous slides) and automatically populated to X(I) column here
- Occasionally the reaction times are not properly read to Arrayed Data Table, and hence not displayed correctly in Data Analysis Panel
- In such a case, click ARR_DATA(I) and the "..." button, define a function of *i* (spectral index) to calculate the reaction times if you can
- Or enter the reaction times manually. You can copy such numbers from a text file and paste to the table





To extract data using the Data Analysis Panel

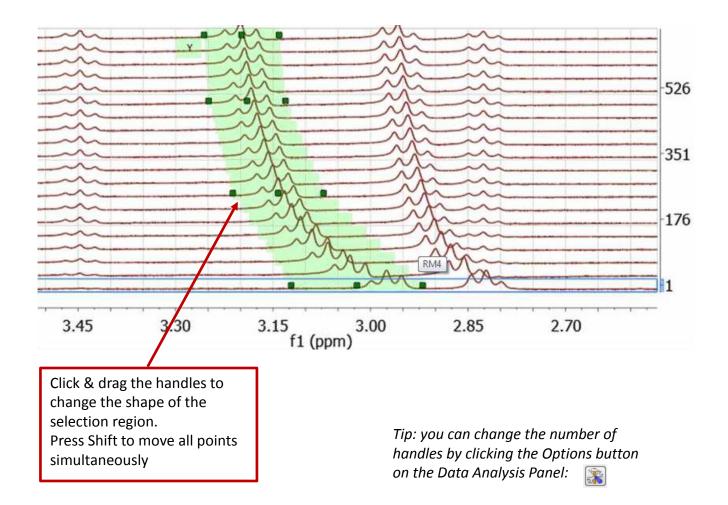
If Integrals is chosen (the default setting), the integrals within the defined region are filled in the Y(X) column, and also plotted in the X-Y graph (X = reaction time, Y= integrals).





To extract data from drifting peaks

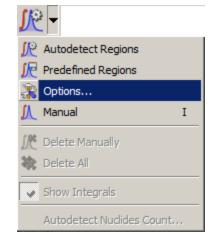
If the peaks drift over time, you can manually change the direction of the integration regions :





More options for peak integration: Sum, GSD/LSD, or Line-fitting methods

- By default, the integration is done by summing up all points within the defined region. This is the default setting – Sum Method.
- When peaks are partially overlapping, it is useful to use the Peaks Method so that only the deconvolved peaks are integrated.
- The is controlled by the Integration Options



👫 Integration Options	? ×
Integral Calculation	ОК
Calculation Method:	Cancel
Sum	Cancel
Sum Peaks	
Source: Autodetect 💌	
Parameters	
Algorithm: Peak Picking 💌	
Minimum Area: 3.00 %	
Defaults	
Apply Automatic Integration	



Peak integration using GSD (Global Spectral Deconvolution)

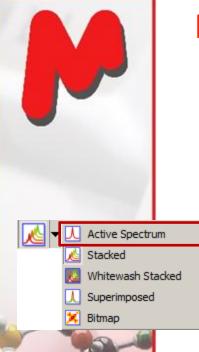
- To use the Peaks-based integration, you can do the following:
 - Do GSD to all the spectra first, and then define the integration region(s), or
 - Do integration directly to the spectra from the Data Analysis Panel: This will do Local Spectral Deconvolution (LSD) to all the spectra within the region*
- You can also use the Line Fitting tools to refine the GSD peaks locally.
- When an integration region is defined from Data Analysis Panel, the deconvolved peaks whose tops fall within the region are summed up as the integral for that spectrum



Peak Picking Options	?
Method:	GSD 💌
Settings	
Refinement Level	Resolution
Ref. 1 (2 fitting cycles)	C High
	C Normal
	• Low
	C Custom 0.25 🛬
Auto Classify	Impurities/Compounds
	Defaults
	OK Cancel

Tip: deconvolution of multiple spectra can be very slow, and unreliable if the peaks are very small. So use it with caution. You are recommended to save the document before such operations.

* LSD can be faster than GSD, but you cannot change the integration region to outside of the current one(s) later, as Mnova will not re-do the LSD if you change the current integration region

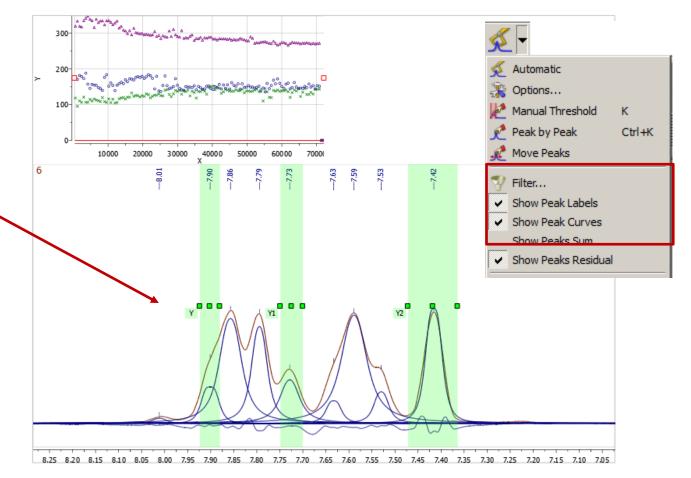


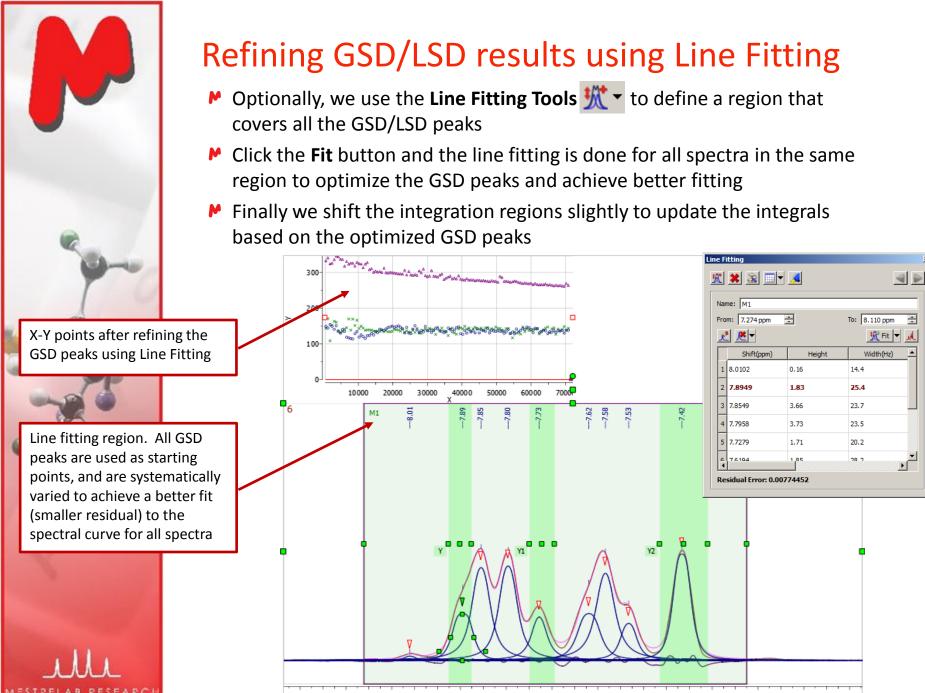
Set to Active Spectrum mode, and turn on the Peak Curves and Residual Curves to see the details of peak deconvolution for each spectrum. Use Shift+ Arrow Keys to browse through the stacked spectra

Chemistry Software Solutions

Peak integration using GSD (Global Spectral Deconvolution)

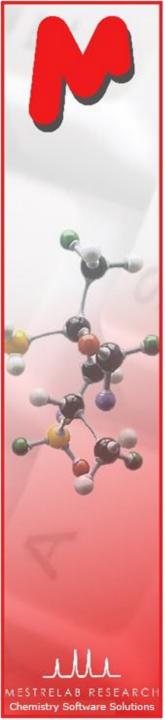
- In this example, LSD is first done within 8.2 7.2 ppm by defining an integration region of 8.2-7.2 ppm
- Next the Integration region is shrunk to cover only the peak around 7.9 ppm, and two more regions are added to cover the other two interesting peaks (7.73 and 7.41 ppm). The details are examined as the following:





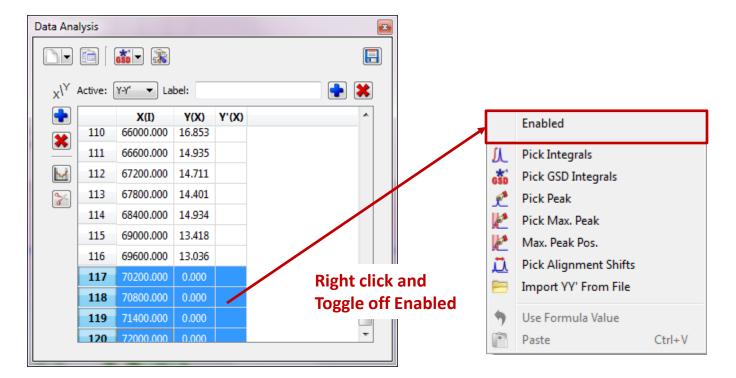
Chemistry Software Solutions

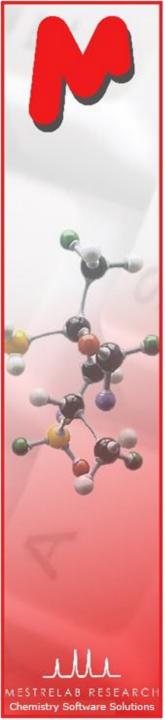
8.15 8.10 8.05 8.00 7.95 7.90 7.85 7.80 7.75 7.70 765 7.60 7.55 7.50 7.45 7.40 7.35 7.30 7.25 7.20



What to do with bad points?

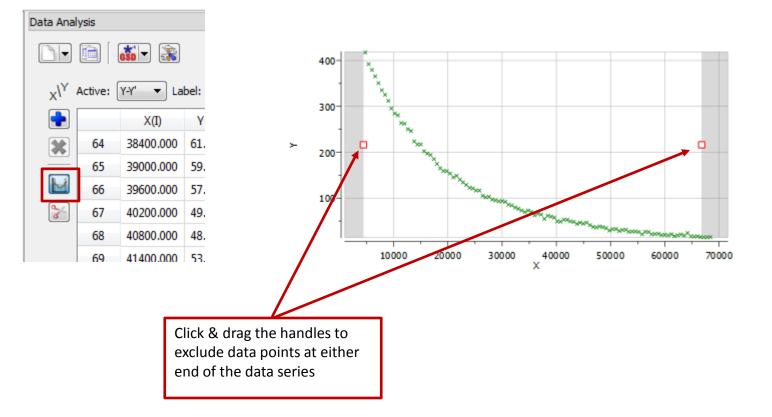
To exclude some data points, highlight them in the table, and right click and turn off Enabled:

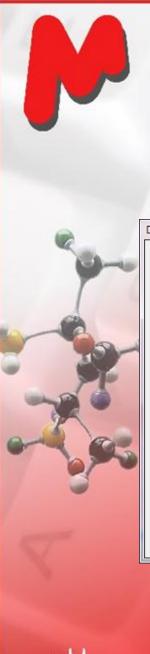




What to do with bad points?

To exclude some data points at the beginning or at the end of the reaction, you can toggle on the Use Fitting Limit button, and exclude the data points on the XY Graph:

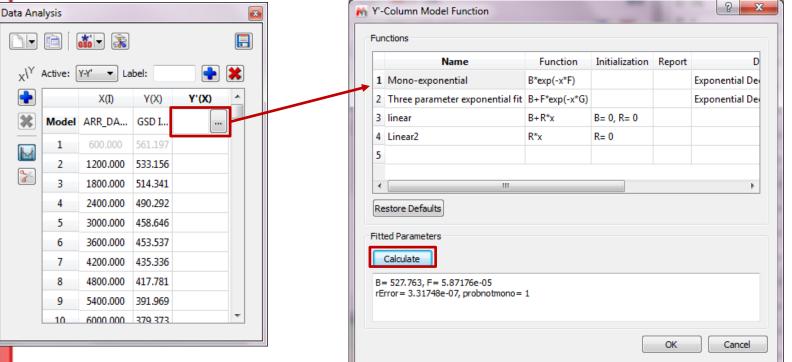




Chemistry Software Solutions

To fit the data to a function

To fit the XY points to a function, double click the first cell in the Y'(X) column, and choose (or define) a function, and click Calculate to do the fitting. Click OK to accept the results:

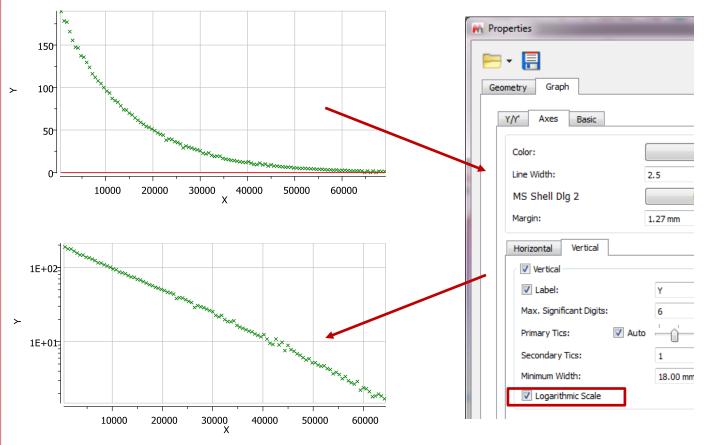


This example shows a first order reaction. F is the rate constant (k). The half-life $t_{1/2} = 0.693/F$



To fit the data to a function

To change the Y scale of the XY graph to logarithmic, right click on the graph and choose Properties, and toggle on the Logarithmic Scale option:

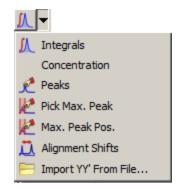


Tip: We recommend you to fit the original values to exponential function directly to avoid bigger numeric fitting errors. Also: Make sure you exclude zero or negative values before converting them to logarithmic scale.



More about the Data Analysis Panel

- You can click the "+" button to add another Y(X) column to the current table/plot, or you can start with a new table/plot by clicking the Create New Plot button.
- You can use other types of spectral properties as Y(X) values:
 - Integrals: peak integrals
 - Concentration: concentrations of an analyte
 - Peaks: intensities of the peaks near a defined location
 - Maximum Peaks: intensities of the highest peaks in a defined region
 - Max. Peak Positions: positions of the highest peaks in a defined region.
 - Pick Alignment Shifts: the shifts of peaks relative to the peak in the first spectrum





Summary

- Mnova NMR provides powerful and easy-to-use tools for processing and analysis of multiple NMR spectra for reaction monitoring
- Such tools can be used for many other types of studies, such as relaxation, diffusion, molecular binding studies, etc.
- See Help | Contents | Advanced Menu | Data Analysis for more info.
- For 45 day free trial of Mnova, go to <u>http://mestrelab.com/software/mnova-suite/download/</u> or email us at sales@mestrelab.com